

mechanical interactions between bacteria and grains in a model soil

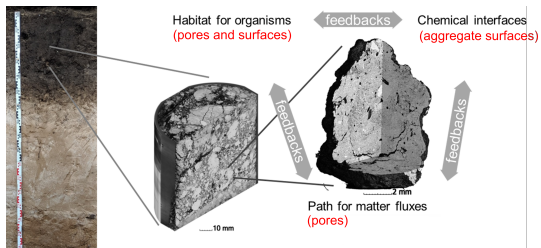
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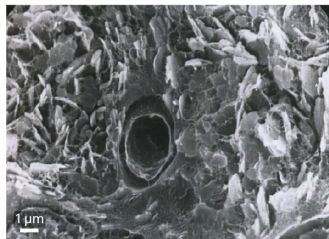
InterPore, Nantes
May 21, 2026



structure ↔ biology feedback in soil



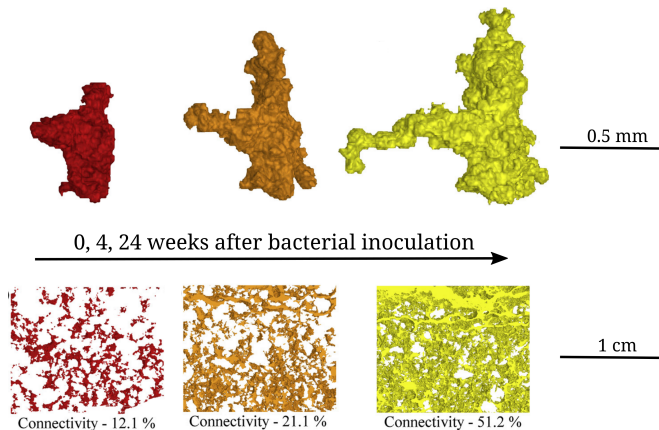
adapted from Schlüter et al. (2020) *Geoderma*



Philipipot et al. (2024) *Nat. Rev. Microbiol.*

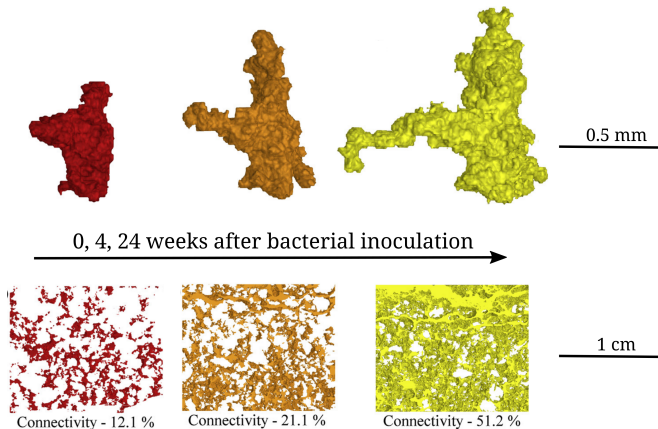
- continuum of pore scales (sub- μm \rightarrow cm)
- biological activity at each scale feeds back with surface chemistry and transport pathways

bacterial inoculation modifies soil porous structure



Heliwell et al. (2014) *Soil Biol Biochem*

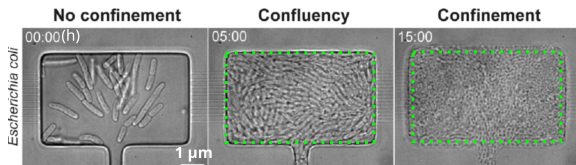
bacterial inoculation modifies soil porous structure



Heliwell et al. (2014) *Soil Biol Biochem*

Bacteria modify pore sizes & increase soil pore connectivity.
Do they mechanically create those spaces?

confined bacteria generate large growth stresses

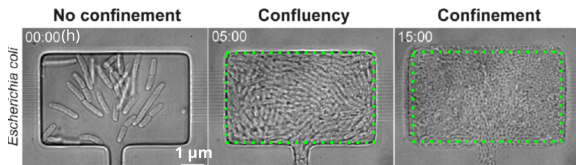


Le Blanc et al. (2024) *bioRxiv*

Turgor
pressure

Internal pressure $>$ turgor pressure

confined bacteria generate large growth stresses



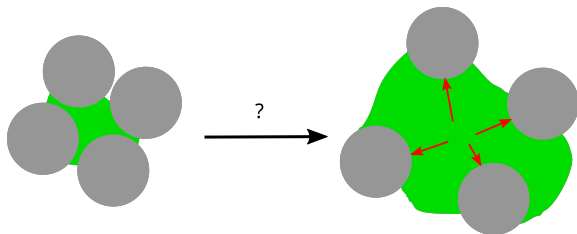
Le Blanc et al. (2024) *bioRxiv*

Turgor
pressure

Internal pressure $>$ turgor pressure

What is the interplay between **growth stresses**, **stress relaxation**, and **cavity deformation** in **divided media**?

research question

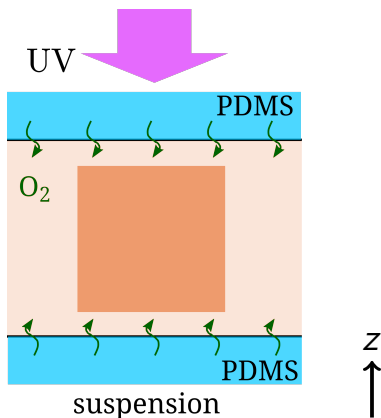


The mechanical action of bacteria on a confining, divided environment has not been directly observed yet.

- Can bacteria deform a granular matrix, and how?
- Which biophysical conditions favour it?

experiments: use **microfluidic techniques** to concurrently observe **bacterial growth** and **position of mobile grains**

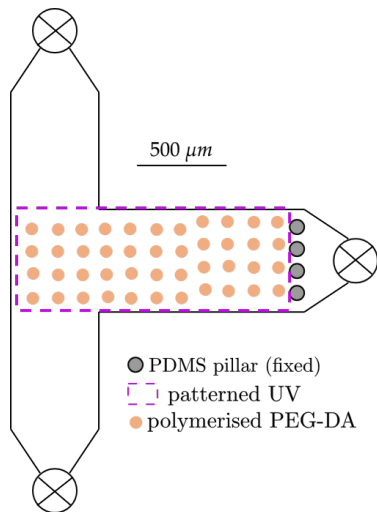
particle fabrication—principle



- **PEG-DA**: hydrogel polymerised (hardened) by UV
- UV projected across microfluidic channel according to pattern

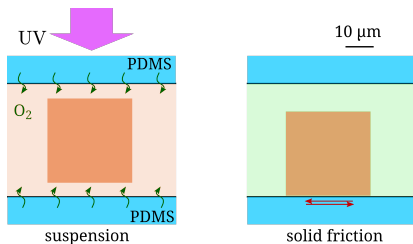
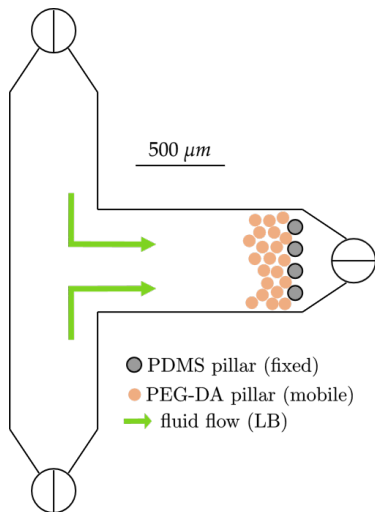
rigid ($\sim 10\text{--}100$ MPa) pillars ($40\ \mu\text{m}$) in suspension

microfluidic chamber—*in situ* particle fabrication



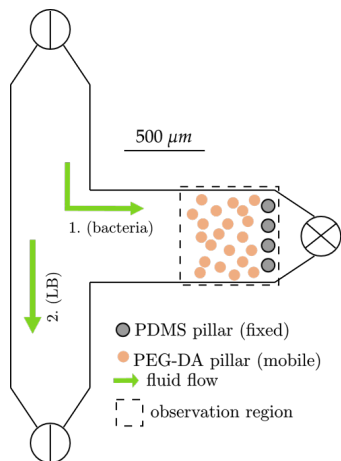
- channel widths $500\ \mu\text{m}$
- channel height $50\ \mu\text{m}$

rendering the environment suitable for bacterial growth



PEG-DA precursor solution rinsed
with 50% spent medium → no
non-specific adhesion

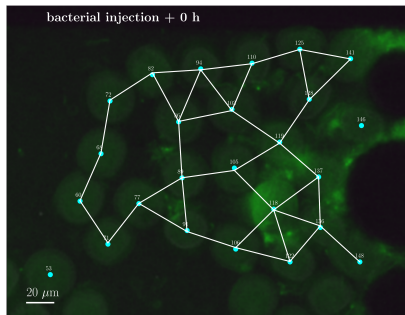
bacterial injection and continuous nutrient supply



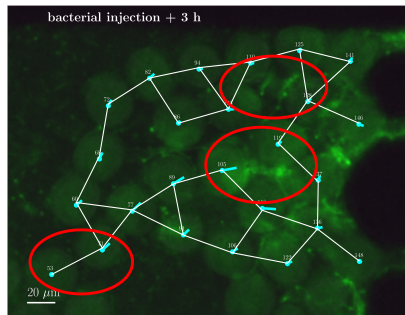
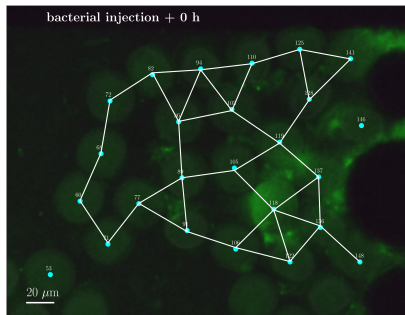
bacteria: *Bacillus subtilis*,
expressing GFP (emit green
when excited by blue)

bacterial growth and pillar movement

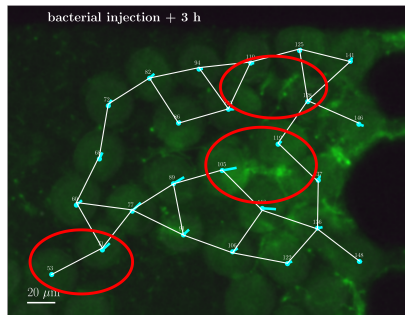
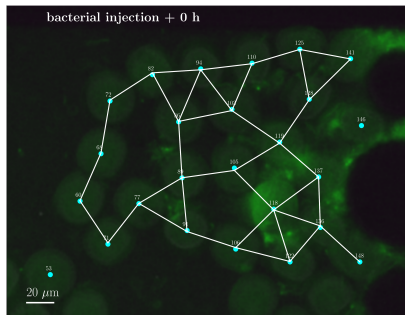
pore widening and movement chains



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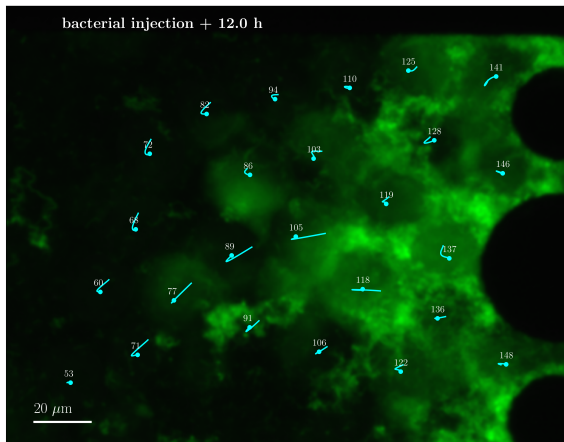
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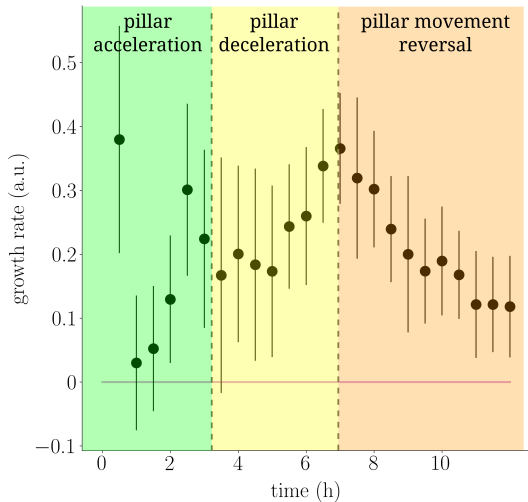
- pillars move when in a bacterial density gradient...
- and when there is free space to move into

idea: bacterial colonies deform soft particle chains

movement reversal

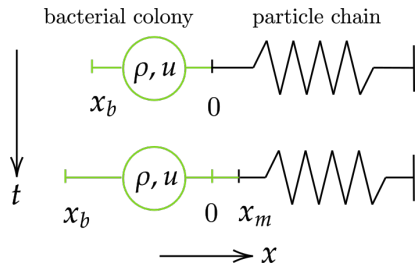


movement reversal is concurrent with growth slowdown

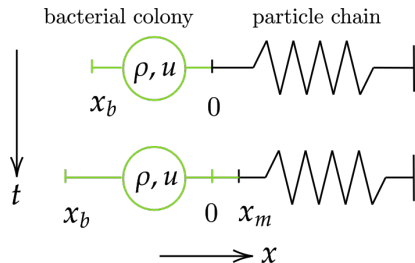


around 7 h after bacterial injection, change in **growth rate**
and in **pillar movement direction**

growth pressure when “partially” confined



growth pressure when “partially” confined



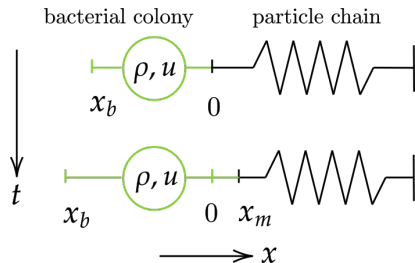
governing equations of bacterial colony:

$$\partial_t \rho + \partial_x (\rho u) = \frac{1}{\tau_g} \rho \quad \text{growth}$$

$$-\partial_x p + \eta \partial_{x^2} u = 0 \quad \text{Stokes flow } (\eta = \eta(\rho)\eta(\phi))$$

$$p = \Pi(\rho), \quad \partial_\rho \Pi > 0 \quad \text{constitutive}$$

growth pressure when “partially” confined



boundary conditions:

$$\rho(x_b) = \eta \partial_x u, \quad \text{free surface}$$

$$\rho(x_m) = kx_m, \quad \text{elastic surface}$$

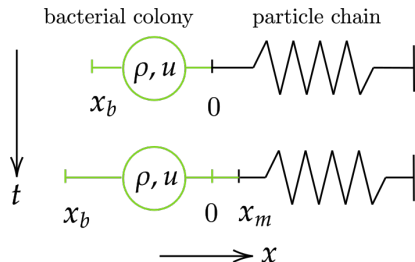
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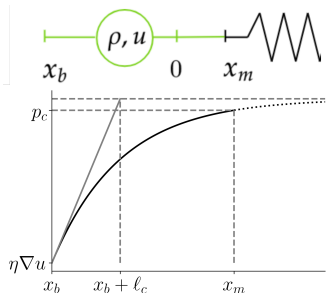
at confluence, $p, u = 0 \Rightarrow$

$$\frac{\partial \rho}{\partial t} > 0 \Rightarrow \frac{\partial p}{\partial t} > 0$$

bacteria generate growth pressure
that drives biomass flow towards the edge of the colony

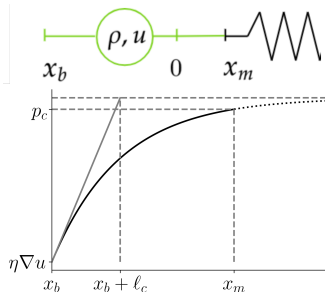
bacterial division vs. advection times

- after confluence, **growth pressure gradient** towards the wall, where $p = p_c$.



bacterial division vs. advection times

- after confluence, **growth pressure gradient** towards the wall, where $p = p_c$.



$$\frac{\partial p}{\partial t} \propto \frac{1}{\tau_g} - \frac{p_c}{\eta}$$

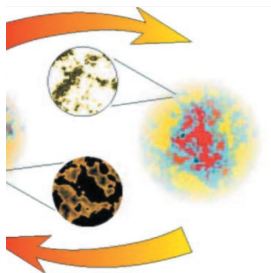
division time
pressure at wall
effective viscosity

- if $\tau_g < \eta/p_c$, bacteria generate pressure faster than they flow, **compressing** the spring $\Rightarrow \frac{\partial p}{\partial t} = k\dot{x}_m > 0$
- if $\tau_g > \eta/p_c$, bacteria flow faster than they divide & the spring **relaxes** $\Rightarrow \frac{\partial p}{\partial t} = k\dot{x}_m < 0$

summary

- pores where bacterial microcolonies were intercepted have **widened** before shrinking back
- toy model: balance growth vs. self-flow \Rightarrow non-monotonous motion
- hypothesis: **fast bacterial growth** favours restructuration by elastically transmitting growth stresses to grains
- current work
 - quantify these stresses
 - understand behaviour at short (1-2 h) times
- perspectives: emerging tension from EPS production? emerging cohesion in the medium?

perspective



Model for self-organization in the soil-microbe communities at a location in soil, and the potential microbial activities, leading to local depletion of oxygen (left-hand concentration represented as in Fig. 3). Increased oxygen changes the local structure, creating a more open structure, which leads to enhanced rates of oxygen supply (right-hand state is used up, activity declines and the structure returns to a closed state. The open and closed states may represent configurations for oxygen supply in a high potential oxygen regime (open) and protection from desiccation and predation in a low oxygen regime (closed).

Young & Crawford (2004) *Science*

acknowledgments



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